













Limestone

Caves Romanian

WISME New Biofilm microbes Cave Thiovulum (Candidatus Thiovulum stygium) differ metabolically and genomically from marine species



NicheVision























| In-Vitro Mixture | Туре | Nanopore Barcode | Total Bases (Mb) | Total Reads | Barcode label |
|------------------|---------------|------------------|------------------|--------------|--------------------------|
| 1 | Single Source | 1 | 383.66 | 89,680 | 1_single_source_17257 |
| 2 | Single Source | 2 | 328.94 | 78, 950 | 2_single_source_17115 |
| 3 | Single Source | 3 | 276.29 | 64,490 | 3_single_source_17066 |
| 4 | Mixed 3:1 | 4 | 440.1 | 103,850 | 4_mixed_3:1_17257v17115 |
| 5 | Mixed 3:1 | 5 | 411.09 | 96,800 | 5_mixed_3:1_17257v17066 |
| 6 | Mixed 3:1 | 6 | 451.16 | 107,050 | 6_mixed_3:1_17115v17066 |
| 7 | Mixed 1:3 | 7 | 435.01 | 103,130 | 7_mixed_1:3_17257v17115 |
| 8 | Mixed 1:1 | 8 | 409.63 | 97,250 | 8_mixed_1:1_17257v17115 |
| 9 | Mixed 10:1 | 9 | 429.52 | 100,750 | 9_mixed_10:1_17257v17115 |
| | Barcode | Primer Forward | Amplicon | Primer Rever | se Barcode |









| Current mtDNA Analysis Workflow |
|--|
| Current practice for mtDNA haplotyping requires single-source samples due to inability to deconvolute mixtures. Being able to deconvolute mtDNA mixtures will enable exploitation of mtDNA from crime scenes. |
| Single Source Sample Sanger or MPS Align to rCRS Reference Call Variants List of Variants |
| Single bone/tooth |















| | | Haplogroup | | Expected Hanlotyne ^{1,2} |
|-----------|-----------------------|--------------------------------|-------------|--|
| Accession | Coriell Population | (Estimated by MixtureAceMT) | Tree Span | Variant Calls |
| NA17257 | Caucasian | U5b2b1b | U5 -> Leaf | 750 1507 2620 268.10 215.10 7500 54005 17717 27665 24070 27665 5664 70207 77665 86665 64774 106660 114670 115030 117764 118206 12208 12208 12630 12637 16630 12637 14500 14670 14670 14670 16630 |
| NA17115 | African American | L1b1a3 | L1b -> Leaf | T26 1520 1520 1531 1530 1540 1540 1540 1540 1540 1541 154 1510 1520 1540 1550 1540 1550 1540 1550 1540 1550 1544 1550 1544 1550 1544 1550 1544 1550 1544 1550 1544 1550 1544 1550 1544 1550 1540 154 |
| NA17066 | Central American | C1c6 | C -> Leaf | 730 249-2620 359-2910 209.42 315.12 4892 7500 14380 18884 27060 35234 36084 47150 47850 70387 79844 85844 87010 88606 92000 95400 16280 162804 162700 107200 117164 110104 12810 127057 131656 128130 143140 147040 145804 152266 156877 152064 161534 16225 142804 152266 156877 152054 161534 16225 |

| | Maximum Likelihood Estimate | | | | Number of | | |
|-------------|-----------------------------|-------------------|--------------|--------------|------------------|-------------|------------|
| Sample | Haplog | roup | Proportion | | Reads | Total Reads | Percent |
| | Expected | Called | Expected | Called | Classified | | Classified |
| CAU | - | U5b2b1b | 1.00 | 1.00 | 25,991 | 89,680 | 29% |
| AA | | L1b1a3 | 1.00 | 1.00 | 17,518 | 78,950 | 22% |
| MX | | C1c6 | 1.00 | 1.00 | 19,332 | 64,490 | 30% |
| 3:1 CAU:AA | U5b2b1b L1b1a3 | U5b2b1b L1b1a3 | 0.75 0.25 | 0.82 0.18 | 21,953 5,096 | 103,850 | 26% |
| 3:1 CAU:MX | U5b2b1b C1c6 | U5b2b1b | 0.75 0.25 | 1.00 | 26,898 | 96,800 | 28% |
| 3:1 AA:MX | L1b1a3 C1c6 | L1b1a3 C1c6 | 0.75 0.25 | 0.61 0.39 | 14,536 9,418 | 107,050 | 22% |
| 1:3 CAU:AA | U5b2b1b L1b1a3 | U5b2b1b L1b1a3 | 0.25 0.75 | 0.27 0.73 | 17,013 6,549 | 103,130 | 23% |
| 1:1 CAU:AA | U5b2b1b L1b1a3 | U5b2b1b L1b1a3 | 0.50 0.50 | 0.56 0.44 | 14,095 11,214 | 97,250 | 26% |
| 10:1 CAU:AA | U5b2b1b | U5b2b1b | 0.91 | 0.94 | 27,255 | 100,750 | 29% |





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Conclusions

 Long range sequencing by Oxford Nanopore Technologies (ONT) combined with probabilistic deconvolution holds promise for mtDNA analysis of mixed samples.

 Here, the mtDNA genome was covered by 6 amplicons to accommodate slightly degraded DNA. However, entire mtDNA chromosomes can possibly be sequenced. If done on mixed samples, the mixture will be "deconvoluted by sequencing", and the haplotypes will be "phased by sequencing".



